

Predictive Value of HIV-1 Genotypic Resistance Test Interpretation Algorithms

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Background. Interpreting human immunodeficiency virus type 1 (HIV-1) genotypic drug-resistance test results is challenging for clinicians treating HIV-1–infected patients. Multiple drug-resistance interpretation algorithms have been developed, but their predictive value has rarely been evaluated using contemporary clinical data sets.

Methods. We examined the predictive value of 4 algorithms at predicting virologic response (VR) during 734 treatment-change episodes (TCEs). VR was defined as attaining plasma HIV-1 RNA levels below the limit of quantification. Drug-specific genotypic susceptibility scores (GSSs) were calculated by applying each algorithm to the baseline genotype. Weighted GSSs were calculated by multiplying drug-specific GSSs by antiretroviral (ARV) potency factors. Regimen-specific GSSs (rGSSs) were calculated by adding unweighted or weighted drug-specific GSSs for each salvage therapy ARV. The predictive value of rGSSs were estimated by use of multivariate logistic regression.

Results. Of 734 TCEs, 475 (65%) were associated with VR. The rGSSs for the 4 algorithms were the variables most strongly predictive of VR. The adjusted rGSS odds ratios ranged from 1.6 to 2.2 ($P < .001$). Using 10-fold cross-validation, the averaged area under the receiver operating characteristic curve for all algorithms increased from 0.76 with unweighted rGSSs to 0.80 with weighted rGSSs.

Conclusions. Unweighted and weighted rGSSs of 4 genotypic resistance algorithms were the strongest independent predictors of VR. Optimizing ARV weighting may further improve VR predictions.

Retrospective studies have shown that the presence of human immunodeficiency virus type 1 (HIV-1) drug resistance before the start of a new antiretroviral (ARV) regimen is an independent predictor of the virologic response (VR) to that regimen [1]. Prospective con-

trolled studies have shown that patients whose physicians have access to drug-resistance data respond better to therapy than those whose physicians do not [2–5]. The accumulation of such retrospective and prospective data has led to the routine use of genotypic resistance testing in the management of HIV-1–infected patients [6–8].

However, interpreting the results of HIV-1 genotypic drug-resistance tests is one of the most difficult challenges facing clinicians who care for HIV-1–infected patients. First, there are many mutations associated with drug resistance [9]. Second, there are synergistic and antagonistic interactions among these drug-resistance mutations [10]. Third, some mutations may not reduce susceptibility by themselves but may compensate for the effect of other mutations [11] or may be sentinel indicators of emerging drug resistance. As a result, several different algorithms have been developed for interpreting HIV-1 genotypic drug-resistance test results.

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Several studies have compared the predictive ability of different genotypic drug-resistance algorithms using retrospective clinical data sets [12–17]. In the present study, we evaluate the predictive value of 4 genotypic drug-resistance test interpretation algorithms in a patient population undergoing a wide range of salvage ARV therapy.

METHODS

Study Subjects and Treatment-Change Episodes (TCEs)

The study subjects were from 16 clinics of the Kaiser-Permanente Medical Care Program–Northern California and had plasma HIV-1 samples sent to Stanford University Hospital for genotypic drug-resistance testing between 1998 and 2007. Eligible subjects included those with a valid TCE, defined as a change in ARVs within 24 weeks of a genotypic drug-resistance test and characterized by the administration of a new salvage regimen for ≥ 4 weeks (figure 1). Additional requirements included a plasma HIV-1 RNA level of >1000 copies/mL obtained within 8 weeks before the ARV change and ≥ 2 plasma HIV-1 RNA levels obtained between 4 and 36 weeks after the ARV change. We also required that a complete list of all the ARVs that a patient had ever received be available.

This protocol received expedited approval by institutional review boards at Stanford University and Kaiser-Permanente Medical Care Program–Northern California. A waiver of consent was provided because only anonymized retrospective data were used.

The VR to the new salvage regimen was classified as sustained for patients with ≥ 2 subsequent plasma HIV-1 RNA levels below the limit of quantification (i.e., <75 copies/mL [hereafter “BLQ”], as determined by use of the Versant bDNA assay [Bayer]), transient for those with a single plasma HIV-1 RNA

level BLQ and absent for those without a plasma HIV-1 RNA level BLQ.

Genotypic Susceptibility Scores (GSSs)

Four genotypic interpretation algorithms were used, including Agence National de Recherche sur le SIDA (hereafter, “ANRS”; version 2007.10) [18], HIV Drug Resistance Database (hereafter, “HIVdb”; version 4.3.1) [19], Rega (version 7.1.1) [20], and ViroSeq (version 2.8) [21]. A 3-step procedure was used to derive a regimen-specific GSS (rGSS) from the baseline genotype (figure 2). First, for each algorithm, the drug-specific GSS for each ARV was calculated. The assigned drug-specific GSS was 1.0 for ARVs for which HIV-1 was predicted to be fully susceptible and 0 for ARVs for which HIV-1 was predicted to be fully resistant. A drug-specific GSS between 0 and 1.0 was assigned to ARVs predicted to have intermediate levels of resistance. For each algorithm, the assigned drug-specific GSS was proportional to the extent of predicted resistance. A drug-specific GSS of 1.0 was assigned to the fusion inhibitor enfuvirtide if it had not been used previously; otherwise, a GSS of 0 was assigned. Second, because ARVs vary in their potency, defined as their antiviral activity and genetic barrier to resistance, we also calculated 2 weighted GSSs. The first weighting entailed multiplying the drug-specific GSSs for boosted protease inhibitors (PIs) by 1.5 (“boosted PI” weighting). This was motivated by the Rega algorithm, the only algorithm of the 4 that provides instructions for creating a weighted GSS. The second “comprehensive” weighting entailed multiplying the drug-specific GSSs for boosted PIs by 2.0 and those for nucleoside (nucleotide) reverse-transcriptase (RT) inhibitors (NRTIs) by 0.5. The GSSs for nonnucleoside RT inhibitors (NNRTIs), enfuvirtide, and nelfinavir were not weighted. Third, the drug-

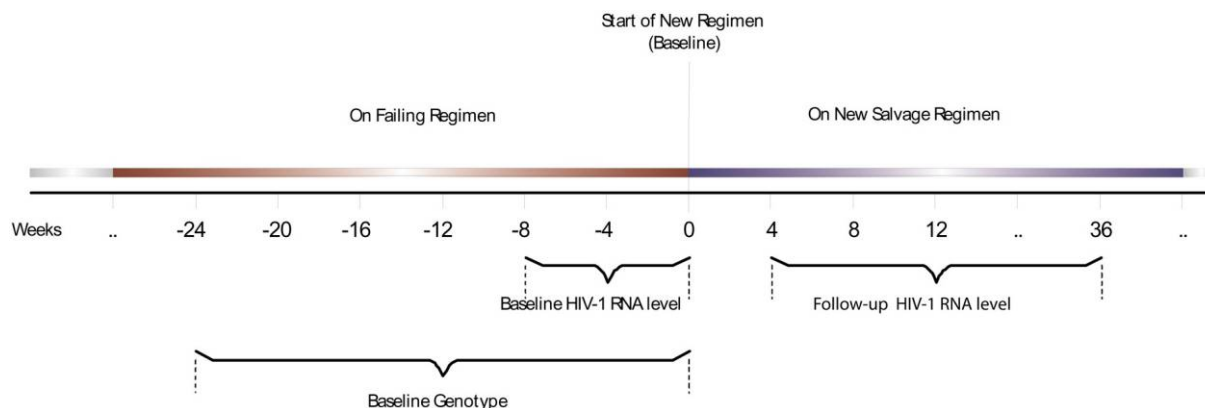


Figure 1. Schematic definition of a treatment-change episode (TCE). Brackets indicate 3 TCE requirements: (1) a baseline genotypic drug-resistance test result obtained within 24 weeks before the treatment change, (2) a baseline plasma human immunodeficiency virus type 1 (HIV-1) RNA level obtained within 8 weeks before the treatment change, and (3) ≥ 2 plasma HIV-1 RNA levels obtained between 4 and 36 weeks after the treatment change. Additional requirements were a CD4 cell count obtained within 24 weeks before the treatment change and ≥ 4 weeks of salvage therapy.



Figure 2. Three-step method for calculating regimen-specific genotypic susceptibility scores (GSSs): (1) calculate drug-specific GSSs, applying a genotypic drug-resistance interpretation algorithm to the mutations present in the baseline genotype to estimate the activity of an antiretroviral (ARV) relative to its activity against a wild-type virus; (2) calculate the weighted drug-specific GSSs, multiplying drug-specific GSSs by a factor that accounts for differences in ARV activity between ARVs belonging to different ARV classes; and (3) calculate regimen-specific GSSs by adding the weighted drug-specific GSSs of the drugs used for salvage therapy.

specific GSSs (whether weighted or unweighted) for each ARV in a regimen were added to obtain an rGSS. We evaluated each of 12 rGSSs (4 algorithms times 3 weighting schemes) in subsequent analyses.

Among patients who received PIs, only those who received nelfinavir or a ritonavir-boosted PI were included because the HIVdb and ViroSeq algorithms provide interpretations for ritonavir-boosted PIs but not unboosted PIs. TCEs from patients who received a combination of ≥ 2 PIs plus ritonavir (dual-boosted PIs) were excluded. Among patients who received NNRTIs, only those who received efavirenz or nevirapine were included, because ANRS does not provide an interpretation for delavirdine. TCEs for which the new salvage regimen contained raltegravir, maraviroc, and etravirine were excluded because these ARVs were licensed only in the last months of the study.

Statistical Analysis

Potential explanatory variables. For each TCE, the following groups of covariates were analyzed, in addition to the rGSS: (1) age, sex, and race; (2) features of the ARV treatment history before the TCE, including duration of therapy, the number of ARVs received, the number of ARV regimens and classes received, the number of PIs received, the number of NRTIs and NNRTIs received, the number of non-highly active ARV therapy (HAART) regimens received, the year ARV therapy was first begun, and the patient's history of ever having attained a VR; (3) plasma HIV-1 RNA level and CD4 cell count at baseline; and (4) features of the salvage regimen, including the year of the TCE and the number of ARVs, new ARVs, and new ARV drug classes received.

Model selection. Univariate analyses were performed to quantify the association between each covariate and VR. Then, for each of the 4 algorithms and 3 GSS weighting schemes (12 models), we performed forward stepwise logistic regression with all covariates that had a statistically significant association with VR in univariate analyses. Among the covariates selected by this approach, those with statistically significant associations ($P < .05$) in ≥ 6 of 12 models were considered robust covariates.

Multivariate logistic regression analyses. We performed

multivariate logistic regression to evaluate the association between each of 12 rGSSs and VR, adjusted for the robust covariates. The adjusted odds ratios (ORs) for attaining VR for each unit increase in the rGSS and the goodness of fit (1–residual deviance/null deviance) were calculated. The prediction of each multivariate logistic regression model was tested using 10-fold cross-validation. With the R package ROCR (version 1.0-2), the receiver operating characteristic (ROC) curve of each prediction result for the 10 test sets was calculated to analyze the trade-off between the proportion of true-positive (correct VR prediction) and false-positive (incorrect VR prediction) results across the range of possible prediction cutoffs [22].

Classification tree. To examine the effects of rGSS and the robust covariates on the numbers of patients who attained VR, we built classification trees for each of the 12 models, using the R package Rpart (Recursive partitioning; version 3.1-39) (<http://cran.r-project.org/web/packages/rpart/index.html>). Tree pruning was performed using the default parameters in Rpart to minimize the complexity of the tree as a function of improvement in classification accuracy.

RESULTS

TCE characteristics. Of the 4654 patients who underwent genotypic drug-resistance testing, 3600 had treatment histories available, including 3029 who received ARVs and 571 who received no ARVs. Of the 3029 ARV-treated patients, 641 patients had 734 valid TCEs (figure 3 and the appendix). The median duration between the baseline genotype and the ARV change was 3 weeks (interquartile range, 1–8 weeks). A mean of 3.4 plasma HIV-1 RNA level test results were available during the 36-week follow-up period, including ≥ 1 plasma HIV-1 RNA level test result available for weeks 1–12, 13–24, and 25–36 for 82%, 80%, and 68% of TCEs, respectively.

The mean age of the patients was 45 years (range, 18–77 years). Ninety-two percent were male; 58% were white, 17% were African American, 16% were Latino, 5% were Asian, and 4% were of unknown race. All but 9 patients were infected with subtype B viruses. The median year in which the TCE occurred was 2002 (range, 1998–2007). The mean baseline

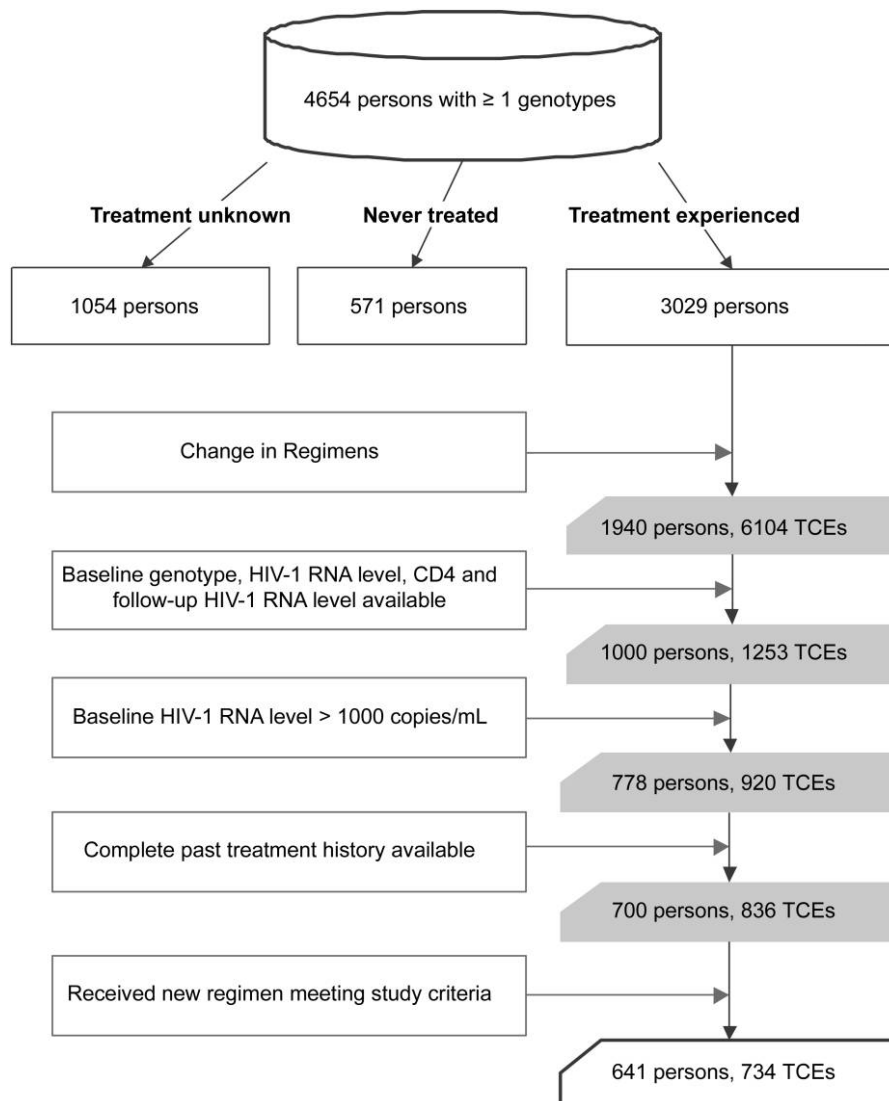


Figure 3. Description of the complete patient population from which the treatment-change episodes (TCEs) were derived. Of 4654 patients who underwent genotypic drug-resistance testing between 1998 and 2007, 3600 patients had a known history of antiretroviral (ARV) treatment. Of these, 3029 had received ≥ 1 ARV drug treatment regimens, and 641 subsequently met study enrollment criteria. These 641 patients had 734 valid TCEs. HIV, human immunodeficiency virus.

plasma HIV-1 RNA level was 4.2 log copies/mL (range, 3.1 to >5.9 log copies/mL), and the mean CD4 cell count was 280 cells/ μ L (range, 3–1110 cells/ μ L). Before the TCE, patients had received ARV therapy for a mean of 5.1 years (range, 1–14 years). Previous ARVs included a mean of 3.5 NRTIs (range, 0–8 NRTIs), 1.7 PIs (range, 0–8 PIs), and 0.6 NNRTIs (range, 0–3 NNRTIs).

Table 1 summarizes the frequency with which different ARVs or ARV combinations were used for salvage therapy. A PI was used in 555 TCEs and an NNRTI in 330 TCEs. One or more NRTIs were used in all but 11 TCEs; the distribution was as follows: 1 NRTI was used in 115 TCEs, 2 in 380 TCEs, 3 in 205 TCEs, and 4 in 22 TCEs. The salvage regimens included

NRTIs and a PI for 374 TCEs (51.0%), NRTIs and an NNRTI for 149 TCEs (20.3%), NRTIs and a PI plus an NNRTI for 170 TCEs (23.2%), and NRTIs only for 30 TCEs (4.1%). Twenty patients received enfuvirtide.

rGSSs by algorithm. The mean unweighted rGSSs for the ANRS, HIVdb, Rega, and ViroSeq algorithms ranged from 1.9 to 2.3; boosted PI-weighted rGSSs ranged from 2.1 to 2.5, and comprehensively weighted rGSSs ranged from 1.8 to 2.2. All 4 algorithms were correlated with R^2 values of 0.71–0.88 for the unweighted rGSSs. However, statistically significant pairwise differences were observed between the rGSSs of the 4 algorithms (data not shown).

For 278 of 734 TCEs, ≥ 1 historical genotype was available

Table 1. Summary of the Most Commonly Used Antiretrovirals (ARVs) for Salvage Therapy

ARV class and regimen
Protease inhibitor ^a
NFV (65)
fAPV/r (69)
ATV/r (55)
IDV/r (69)
LPV/r (183)
SQV/r (94)
TPV/r (12)
DRV/r (8)
Nucleoside reverse-transcriptase inhibitor
ddl + AZT/d4T (93)
3TC/FTC + AZT/d4T (84)
TDF + 3TC/FTC (56)
AZT/d4T (54)
TDF + 3TC/FTC + AZT/d4T (53)
ABC + 3TC/FTC + AZT/d4T (47)
ABC + AZT/d4T (44)
TDF + ddl + 3TC/FTC (40)
ABC (27)
ddl + 3TC/FTC + AZT/d4T (26)
Nonnucleoside reverse-transcriptase inhibitor
EFV (235)
NVP (95)
Fusion inhibitors
Enfuvirtide (20)

NOTE. No. in parentheses indicates the numbers of treatment-change episodes in which the ARV or ARV combination was included in the salvage therapy regimen. 3TC, lamivudine; 3TC/FTC, 3TC or emtricitabine; ABC, abacavir; ATV, atazanavir; AZT, zidovudine; AZT/d4T, AZT or stavudine; ddl, didanosine; DRV, darunavir; EFV, efavirenz; fAPV, fosamprenavir or amprenavir; IDV, indinavir; LPV, lopinavir; NFV, nelfinavir; NVP, nevirapine; SQV, saquinavir; TDF, tenofovir; TPV, tipranavir.

^a Ritonavir boosting is denoted by “/r.”

before the baseline genotype, including 1 genotype for 168 TCEs and ≥ 2 genotypes for 110 TCEs. For all 4 algorithms, the mean unweighted rGSS calculated using the baseline genotype (range, 1.9–2.4) was higher than the mean unweighted rGSS based on a cumulative genotype containing the union of mutations in the baseline and historical genotypes (range, 1.6–2.0; $P < .001$).

VR and TCEs. Sixty-five percent of the TCEs (475 of 734) were associated with VR, including 51% ($n = 378$) with a sustained VR, defined as ≥ 2 plasma HIV-1 RNA measurements BLQ within the first 36 weeks of follow-up, and 13% ($n = 97$) with a transient VR, defined as a single plasma HIV-1 RNA measurement BLQ. Multivariate logistic regression yielded a slightly better goodness of fit when the transient responders were grouped with the sustained responders rather than with the nonresponders (data not shown); therefore, for the analyses that follow, the sustained and transient responders were grouped together. There was no difference between the good-

ness of fit of models, including all 734 TCEs from 641 patients (77 patients had >1 TCE), and that of models, including only 1 TCE per patient (data not shown). Therefore, for the analyses that follow, we included all 734 TCEs.

Univariate analyses of predictors of VR. Table 2 summarizes the univariate analyses of the association between VR and each of the unweighted rGSSs and the covariates. For each algorithm, a higher rGSS was associated with a greater likelihood of VR. Among the covariates related to past treatment, the number of ARVs and ARV regimens previously received, the number of PIs and ARV classes previously received, and the number of non-HAART regimens received were associated with a decreased likelihood of VR. Among the covariates related to the salvage regimen, the number of ARVs received, the number of new ARVs received, and the number of new ARV classes received were associated with increased VR. A lower baseline plasma HIV-1 RNA level, a higher baseline CD4 cell count, a history of previous VR, and a more recent TCE were also associated with increased VR.

Multivariate logistic regression. Twelve multivariate logistic regression models were developed, 1 for each combination of the 4 algorithms and 3 weighting schemes. In forward selection procedures for all 12 models, there was a statistically significant association between rGSS and VR ($P < .001$). Four covariates were robustly associated with VR—that is, they were significantly associated with VR in ≥ 6 of the 12 models. The robust covariates were as follows: (1) baseline plasma HIV-1 RNA level (12 models), (2) history of previous virologic suppression (12 models), (3) number of new ARVs received (12 models), and (4) number of new ARV classes received (6 models).

The adjusted ORs for the rGSSs demonstrated an increased likelihood of VR, with a range of 63%–71% for each 1.0 increase in unweighted rGSS (i.e., the addition of 1 fully active ARV), 81%–89% for each 1.0 increase in the boosted PI-weighted rGSSs (i.e., the addition of a boosted PI predicted to retain two-thirds of its activity or any other fully active ARV), and 108%–124% for each 1.0 increase in the comprehensively weighted rGSS (i.e., the addition of a boosted PI that retained half of its activity, 2 fully active NRTIs, or any other fully active ARV) (table 3).

We assessed the predictive accuracy of each model by measuring the area under the ROC curve (AUC) generated by 10-fold cross-validation testing. For each weighting scheme, the averaged AUCs of 4 multivariate models (1 for each of the 4 algorithms) were nearly identical (table 3). When the unweighted rGSS was used, the averaged AUC was 0.76 for all 4 algorithms. However, when the comprehensively weighted rGSS was used, the averaged AUC was 0.80 for 3 algorithms and 0.79 for the fourth (ANRS) (table 3).

Multivariate models for unweighted GSSs for all 4 algorithms

Table 2. Univariate Analyses of Virologic Response to Treatment-Change Episodes as a Function of Demographic Factors, Baseline Disease Factors, and Salvage Therapy

Variable	Nonresponders (n = 259)	Responders (n = 475)	P ^a
Characteristic of treatment history			
ARV therapy, years	5 (3–7)	5 (3–7)	.3
ARVs, no.	6 (4–8)	5 (3–7)	<.001
ARV regimens, no.	4 (2–6)	3 (1–4)	<.001
Non-HAART regimens, no.	2 (1–3)	1 (0–3)	<.001
Protease inhibitors, no.	2 (1–3)	1 (1–2)	<.001
NRTIs, no.	4 (2–4)	3 (2–4)	.006
NNRTIs, no.	1 (0–1)	0 (0–1)	<.001
ARV classes, no.	3 (2–3)	2 (2–3)	<.001
Year ARV therapy begun	1996 (1994–1997)	1996 (1994–1998)	<.001
Previous HIV-1 RNA BLQ, %	20.1	41.3	<.001 ^b
Patient characteristic at baseline			
Age, years	42 (38–49)	45 (39–51)	.009
Female sex, %	7.7	8.2	.03 ^b
HIV-1 RNA, log copies/mL	4.4 (4–4.9)	4.1 (3.6–4.6)	<.001
CD4 cell count, cells/mL	190 (94.5–327)	283 (175–407.5)	<.001
Characteristic of salvage regimen			
ARVs, no.	3 (3–4)	3 (3–4)	.01
Previously unused ARVs, no.	1 (1–2)	2 (1–2)	<.001
Previously unused ARV classes, no.	0 (0–1)	1 (0–1)	<.001
Year of treatment-change episode	2000 (1999–2002)	2001 (2000–2003)	<.001
Unweighted rGSS ^c			
ANRS	2 (1–3)	3 (2–3)	<.001
HIVdb	1.5 (0.75–2.25)	2.25 (1.5–2.75)	<.001
Rega	2 (1–2.5)	2.5 (2–3)	<.001
ViroSeq	1.5 (1–2.5)	2.5 (2–3)	<.001

NOTE. Except for categorical data, given as percentages, data are given as medians, with interquartile ranges in parentheses. ANRS, Agence National de Recherche sur le SIDA; ARV, antiretroviral; BLQ, below the limit of quantification (<75 copies/mL; Versant branched DNA assay); HAART, highly active antiretroviral therapy; HIV-1, human immunodeficiency virus type 1; HIVdb, HIV Drug Resistance Database; NRTIs, nucleoside reverse-transcriptase (RT) inhibitors; NNRTIs, nonnucleoside RT inhibitors; rGSS, regimen-specific genotypic susceptibility score.

^a Student's *t* test was used for all comparisons, except as indicated for categorical data.

^b Comparisons by χ^2 test.

^c Algorithm versions were as follows: ANRS 2007.10, HIVdb 4.3.1, Rega 7.1.1, and ViroSeq 2.8.

were highly concordant in predicting VR. Of the 475 TCEs with a VR, when a probability of response of ≥ 0.5 was used 403 of 475 were correctly predicted (85% true-positive), and 47 of 475 were incorrectly predicted (10% false-negative) by all 4 algorithms; only 25 (5%) of 475 yielded discordances among the algorithms. Of the 259 TCEs without a VR, 113 of 259 were correctly predicted (44% true-negative), and 116 of 259 were incorrectly predicted (45% false-positive) by all 4 algorithms; only 30 (11%) of 259 yielded discordances among the algorithms. At a probability-of-response cutoff of 0.8, the false-positive rate decreased from 45% to 10% at the expense of an increased false-negative rate.

Of 734 TCEs, there were 54, 384, and 296 associated with experience with 1, 2, and 3 ARV classes, respectively. Although

the VR rate was higher among patients with experience of 2 classes versus those with experience of 3 classes (74% vs. 53%; $P < .001$), the predictive accuracy was higher among those with experience of 3 classes for all 4 algorithms. For 278 TCEs with ≥ 1 historical genotype, the predictive accuracy did not differ significantly depending on whether a baseline or a cumulative genotype was used.

Classification trees. For all 12 classification trees, the rGSS was the optimal first classifier for VR. Figure 4 shows the tree for the HIVdb algorithm using the comprehensive weighting scheme: VR occurred in 24% of patients (35 of 148) with an rGSS of < 1.1 , compared with 75% of patients (440 of 586) with an rGSS of ≥ 1.1 . Among those with an rGSS of ≥ 1.1 , a further increase in VR was associated with a baseline HIV-1 RNA level

Table 3. Multivariate Logistic Regression Analysis Evaluating the Virologic Function of 3 Weighting Schemes

Algorithm ^a	Mean rGSS ± SD	Adjusted OR of rGSS ^b	Mean AUC ± SD ^c
Unweighted			
ANRS	2.26 ± 0.94	1.70	0.76 ± 0.08
HIVdb	1.91 ± 0.91	1.71	0.76 ± 0.08
Rega	2.22 ± 0.91	1.63	0.76 ± 0.08
ViroSeq	2.14 ± 0.96	1.66	0.76 ± 0.08
Boosted PI weighting			
ANRS	2.51 ± 1.03	1.89	0.77 ± 0.05
HIVdb	2.12 ± 0.99	1.88	0.77 ± 0.05
Rega	2.49 ± 0.91	1.83	0.77 ± 0.06
ViroSeq	2.37 ± 1.05	1.81	0.77 ± 0.05
Comprehensive weighting			
ANRS	2.14 ± 0.97	2.14	0.79 ± 0.03
HIVdb	1.82 ± 0.88	2.22	0.80 ± 0.03
Rega	2.18 ± 0.91	2.15	0.80 ± 0.04
ViroSeq	2.02 ± 0.95	2.08	0.80 ± 0.03

NOTE. For boosted protease inhibitor (PI) weighting, the drug-specific genotypic susceptibility score (GSS) was multiplied by 1.5 before the regimen-specific GSS (rGSS) was calculated. This is the weighting used by the Rega algorithm. For comprehensive weighting, the drug-specific GSS was multiplied by 2.0 for boosted PIs and by 0.5 for nucleoside reverse-transcriptase inhibitors before the rGSS was calculated. ANRS, Agence National de Recherche sur le SIDA; AUC, area under the receiver operating characteristic curve; HIVdb, HIV Drug Resistance Database; OR, odds ratio; SD, standard deviation.

^a Algorithm versions were as follows: ANRS 2007.10, HIVdb 4.3.1, Rega 7.1.1, and ViroSeq 2.8.

^b Adjusted ORs for attaining virologic suppression (plasma human immunodeficiency virus type 1 RNA level below the limit of quantification) for each unit increase in rGSS. For all ORs in this column, *P* values were <.001.

^c Mean AUCs were calculated from the prediction results of multivariate logistic regression models tested in 10-fold cross-validation.

<4.7 log copies/mL, an rGSS ≥1.8, and a history of ever having attained VR. Among those with an rGSS of <1.1, the likelihood of VR was increased in patients receiving ≥1.5 new ARVs who had a history of VR.

rGSSs and VR by ARV class. To assess the relative observed and predicted activity of boosted PIs compared with NNRTIs, we examined the VR to therapy among 119 patients receiving an initial NNRTI-containing salvage regimen (lacking a PI) and 76 patients receiving an initial boosted PI-containing salvage regimen (but lacking an NNRTI), using the unweighted HIVdb algorithm. Among the 119 patients receiving an initial NNRTI-containing salvage regimen, virologic failure occurred in 35%; in contrast, among 76 patients receiving an initial boosted PI-containing salvage regimen, virologic failure occurred in only 12% (*P* < .001). This improved VR with the boosted PI-containing regimen occurred even though the mean rGSSs were almost identical for the boosted PI- and NNRTI-containing regimens (rGSS, ~2.3). However, when comprehensive weighting was used, the rGSS of the boosted PI-containing regimen

was higher than that of the NNRTI-containing regimen (2.5 vs. 1.7; *P* < .001).

DISCUSSION

Rationale for genotypic drug-resistance interpretation algorithms. Genotypic drug-resistance testing is part of the standard care of HIV-1-infected patients. However, because the genetic mechanisms of ARV resistance are both numerous and complex, the mutation list generated by resistance testing has meaning only to a small subset of care providers well versed in the HIV resistance literature. The majority of care providers must rely on the results of a genotypic drug-resistance test interpretation that estimates the extent of ARV resistance associated with the list of reported mutations.

A large number of studies have been performed to discover rules that correlate pretherapy drug-resistance mutations with the clinical response to a salvage therapy ARV. The ARV may be a licensed ARV or a new ARV under clinical investigation. In both cases, the ARV is usually given in combination with a

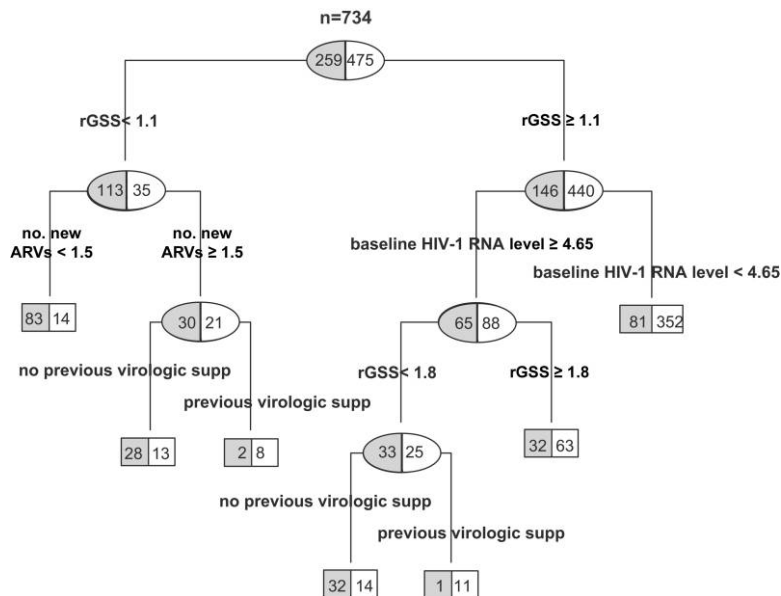


Figure 4. Classification tree for the comprehensively weighted version of regimen-specific genotypic susceptibility scores (rGSSs) generated by the HIVdb algorithm. The root node contains all 734 treatment-change episodes (TCEs). The root node and all child nodes show the number of TCEs resulting in virologic failure (left side of node) or virologic success (right side of node). Using the R package Rpart (Recursive partitioning; version 3.1-39), an explanatory variable and a cutoff value were selected at each node to optimally separate virologic responders from nonresponders in the child nodes. Tree pruning was performed using the default parameters in Rpart for minimizing the complexity of the tree as a function of the improvement in classification accuracy. ARV, antiretroviral; HIV, human immunodeficiency virus.

background regimen that has been optimized for ARV potency. The RESIST, POWER, and DUET studies are examples of studies that have yielded drug-specific rules [23–25]. Indeed, there are >50 published studies of this type, including >30 for PIs and >20 for RT inhibitors [26].

However, rules-discovery studies are limited in power because of the large numbers of drug-resistance mutations and covariates that influence VR. Rules-discovery studies also differ in how VR has been defined and in the spectrum of ARVs available for the optimized background regimen [27]. Therefore, several groups have developed integrated genotypic drug-resistance test interpretation algorithms that supplement rules discovered from clinical studies with other types of information, including in vitro susceptibility testing and in vitro and in vivo drug selection.

Predictive value of genotypic drug-resistance interpretation algorithms. In this study we found that in multivariate logistic regression analyses, rGSSs as determined by 4 interpretation algorithms were the strongest predictors of VR. Indeed, the adjusted OR for the unweighted rGSS of the 4 algorithms ranged from 1.6 to 1.7. In addition, the rGSS was found to be the optimal first classifier for VR in all 12 classification trees (4 algorithms times 3 weightings).

Other statistically significant covariates included the baseline plasma HIV-1 RNA level, the number of new ARVs and ARV classes in the salvage regimen, and a history of previous vi-

rologic suppression. The significance of baseline plasma HIV-1 RNA levels and of the number of new ARVs and new ARV classes as predictors of VR is consistent with the results of previous studies [12, 15, 17]. A history of previous VR may be a surrogate for patient adherence [12].

Although multivariate logistic regression was useful for estimating the predictive value of the algorithms in this study, the model we developed was not designed to be a clinical tool for predicting VR in individual patients. Moreover, although the AUCs of cross-validated ROC curves are robust indicators of prediction accuracy, maximizing AUCs may not be as clinically meaningful as lowering the false-positive rates.

This study was performed before the widespread impact of 3 second-generation drugs (tipranavir, darunavir, and etravirine) and drugs belonging to 2 new drug classes (the integrase inhibitor raltegravir and the chemokine [C-C motif] receptor 5 inhibitor maraviroc). Therefore, the utility of genotypic drug-resistance interpretation algorithms found in this study is particularly relevant to salvage therapy in low- and middle-income countries, where there is less access to the newer, more expensive ARVs that have recently become available in high-income countries.

Weighting of drug-specific GSSs. The finding that the predictive value of all 4 algorithms improved with weighting suggests that weighting GSSs by ARV potency may improve VR prediction. Two findings in particular supported weighting

schemes that attribute increased potency to boosted PIs. First, initial boosted PI-containing salvage regimens lacking an NNRTI were associated with about one-third as many virologic failures (35% vs. 12%) as initial NNRTI-containing salvage regimens lacking a PI. Second, weighting schemes that attributed increased potency to boosted PIs were associated with an increased prediction accuracy for all 4 algorithms. These findings are consistent with those of another recent study showing that PI-specific GSSs were more predictive than NRTI- and NNRTI-specific GSSs [15].

Although it is possible to incorporate ARV potency into a genotypic drug-resistance interpretation algorithm, this is not consistently done. Instead, most algorithms create ARV-specific GSSs that primarily indicate the expected activity of an ARV relative to its activity against a wild-type virus (which is how antimicrobial resistance test results are typically reported). Because ARVs differ in their potency, a second layer of logic may be necessary for choosing among ARVs, particularly when multiple ARVs are predicted to be active. However, our study was not optimal for discovering drug potency weights for individual ARVs, because the number of cases per ARV was insufficient for many ARVs, and the extent to which each algorithm already incorporated ARV potency was unknown.

Conclusion. In this study, both unweighted and weighted rGSSs were the strongest predictors of VR for 4 genotypic drug-resistance interpretation algorithms. Whereas weighting drug-specific GSSs by ARV potency appears to improve predictive accuracy, the weighting schemes we used in this study represent initial steps toward incorporating ARV potency into interpretation of genotypic drug-resistance test results. Ongoing studies of this type are required as new ARVs are licensed and new knowledge about drug-resistance mutations accrues. Further studies are also needed to explore ARV-specific weightings that would more faithfully account for the potencies of individual ARVs and ARV combinations.

APPENDIX

The complete set of data comprising each TCE and the R code used for analysis will be made available at <http://hivdb.stanford.edu> at the time of publication. The baseline genotypes are in GenBank. The accession numbers are listed below.

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